

ANTIBODIES THAT BIND HUMAN INTERLEUKIN-18 AND METHODS OF MAKING AND USING

Related Applications

5 This application is a non-provisional application claiming priority to U.S. provisional application Serial No. 60/181,608, filed February 10, 2000, entitled, "Antibodies that Bind Human Interleukin-18 and Methods of Making and Using" the contents of which are hereby incorporated by reference. In addition, the contents of all cited references, including literature references, issued patents, and published patent
10 applications, as cited throughout this application are hereby expressly incorporated by reference.

Background of the Invention

15 Interleukin-18 (IL-18) was originally described in 1989 as interferon-gamma inducing factor (IGIF) and is a pro-inflammatory cytokine with various functions in addition to an ability to induce interferon gamma. These biological properties include activation of NF- κ b, Fas ligand expression, the induction of both CC and CXC chemokines, and increased production of competent human immunodeficiency virus.

20 Due to the ability of IL-18 to induce interferon gamma production in T cells and macrophages, it plays an important role in Th1-type immune responses and participates in both innate and acquired immunity. IL-18 is related to the IL-1 family in terms of both structure and function. For reviews of IL-18 structure, function and biological activity, see for example Dinarello, C. et al. (1998) *J. Leukoc. Biol.* 63:658-654; Dinarello, C.A. (1999) *Methods* 19:121-132; and Dinarello, C.A. (1999) *J. Allergy Clin.*
25 *Immunol.* 103:11-24.

 It would be desirable to use to modulate IL-18 in a variety of human immune responses. In particular, antibodies that bind to and neutralize IL-18 are particularly desirable. Moreover, murine IL-18 antibodies are limited for their use *in vivo* due to problems associated with administration of mouse antibodies to humans, such as short
30 serum half life, an inability to trigger certain human effector functions and elicitation of an unwanted immune response against the mouse antibody in a human (the "human anti-mouse antibody" (HAMA) reaction).

 In general, attempts to overcome the problems associated with use of fully-murine antibodies in humans, have involved genetically engineering the antibodies to be
35 more "human-like." For example, chimeric antibodies, in which the variable regions of the antibody chains are murine-derived and the constant regions of the antibody chains are human-derived, have been prepared (Junghans, *et al.* (1990) *Cancer Res.* 50:1495-1502; Brown *et al.* (1991) *Proc. Natl. Acad. Sci.* 88:2663-2667; Kettleborough *et al.*

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(1991) *Protein Engineering*. 4:773-783). However, because these chimeric and humanized antibodies still retain some murine sequences, they still may elicit an unwanted immune reaction, the human anti-chimeric antibody (HAMA) reaction, especially when administered for prolonged periods.

- 5 A preferred IL-18 inhibitory agent to murine antibodies or derivatives thereof (e.g., chimeric or humanized antibodies) would be an entirely human anti-IL-18 antibody, since such an agent should not elicit the HAMA reaction, even if used for prolonged periods. However, such antibodies have not been described in the art and, therefore are still needed.

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Summary of the Invention

This invention pertains to compounds, such as antibodies, that bind human IL-18, as well as methods of making and using such compounds or antibodies.

- 15 In one aspect, the invention pertains to a compound capable of binding a human IL-18 amino acid sequence, or portion thereof, where the amino acid comprises an N- or C-terminal portion of human IL-18 such as provided in SEQ ID NO: 70 or SEQ ID NO: 71. In one embodiment, the compound is a small molecule, peptide, polypeptide, antibody, or antibody fragment, such as a fully human antibody or fragment.

- 20 In another aspect, the invention pertains to a human monoclonal antibody, or antigen-binding portion thereof, capable of binding to human IL-18. In other embodiments, the antibody or fragment thereof, dissociates from human IL-18, as determined by plasmon resonance, with a k_{off} rate constant of 0.1 s⁻¹ or less, 1 x 10E-2 s⁻¹ or less, 1 x 10E-3 s⁻¹ or less, 1 x 10E-4 s⁻¹ or less, 1 x 10E-5 s⁻¹ or less, 1 x 10E-6 s⁻¹ or less, or inhibits human IL-18 activity with an IC₅₀ of 1 x 10E-6 or less, 1 x 10E-7 or less, 1 x 10E-8 or less, 1 x 10E-9, 1 x 10E-10 or less, or 1 x 10E-11 or less.

- 25 In another aspect, the invention pertains to an isolated antibody, or an antigen-binding portion thereof, that binds an epitope of human IL-18 comprising amino acids PLFEDMTDSDCRDNA (SEQ ID NO: 1), VIRNLNDQVLFIDQ (SEQ ID NO: 33), or a portion of either. Preferably, the antibody is a neutralizing antibody. Preferably, the antibody is a human antibody. In various embodiments, the antibody is a recombinant antibody (e.g., a single-chain antibody (scFv)), or a monoclonal antibody.

- 30 In other embodiments, the isolated antibody, or antigen-binding portion thereof, binds to an epitope of human IL-18, or a portion of either, where the antibody, or antigen-binding portion thereof, dissociates from human IL-18 with a k_{off} rate constant of 0.1 s⁻¹ or less, as determined by surface plasmon resonance, or which inhibits human IL-18 activity with an IC₅₀ of 1 x 10⁻⁶ M or less. Alternatively, the antibody, or an antigen-binding portion thereof, may dissociate from human IL-18 with a k_{off} rate constant of 1 x 10⁻² s⁻¹ or less, as determined by surface plasmon resonance, or may

- inhibit human IL-18 activity with an IC_{50} of $1 \times 10^{-7}M$ or less. Alternatively, the antibody, or an antigen-binding portion thereof, may dissociate from human IL-18 with a k_{off} rate constant of $1 \times 10^{-3}s^{-1}$ or less, as determined by surface plasmon resonance, or may inhibit human IL-18 activity with an IC_{50} of $1 \times 10^{-8}M$ or less. Alternatively, the
- 5 antibody, or an antigen-binding portion thereof, may dissociate from human IL-18 with a k_{off} rate constant of $1 \times 10^{-4}s^{-1}$ or less, as determined by surface plasmon resonance, or may inhibit human IL-18 activity with an IC_{50} of $1 \times 10^{-9}M$ or less. Alternatively, the antibody, or an antigen-binding portion thereof, may dissociate from human IL-18 with a k_{off} rate constant of $1 \times 10^{-5}s^{-1}$ or less, as determined by surface plasmon resonance, or
- 10 may inhibit human IL-18 activity with an IC_{50} of $1 \times 10^{-10}M$ or less. Alternatively, the antibody, or an antigen-binding portion thereof, may dissociate from human IL-18 with a k_{off} rate constant of $1 \times 10^{-6}s^{-1}$ or less, as determined by surface plasmon resonance, or may inhibit human IL-18 activity with an IC_{50} of $1 \times 10^{-11}M$ or less.

- Another aspect of the invention pertains to an isolated human antibody, or an
- 15 antigen-binding portion thereof, containing at least one variable region CDR domain capable of binding an epitope of human IL-18. In related embodiments, the isolated antibody, or an antigen-binding portion thereof, has a variable region containing a heavy and/or light chain CDR1 domain, CDR2 domain, or CDR3 domain as set forth in Table 6 or 9 which can have, *e.g.*, one or more amino acid substitutions or insertions at or
- 20 adjacent to any of the Kabat positions indicated in Tables 7-8 and 10-11. In a preferred embodiment, the isolated antibody, or an antigen-binding portion thereof, contains a light chain variable region (LCVR) containing the amino acid sequence of SEQ ID NO: 29 and a heavy chain variable region (HCVR) containing the amino acid sequence of
- 25 SEQ ID NO: 26. In another preferred embodiment, the isolated antibody, or an antigen-binding portion thereof, contains a light chain variable region (LCVR) having the amino acid sequence of SEQ ID NO: 29 and a heavy chain variable region (HCVR) having the amino acid sequence of SEQ ID NO: 27.

- Another aspect of the invention pertains to pharmaceutical compositions comprising an antibody, or antigen-binding portion thereof, of the invention and a
- 30 pharmaceutically acceptable carrier. In one embodiment, the pharmaceutical composition further comprises at least one additional therapeutic agent for treating a disorder in which IL-18 activity is detrimental.

- Another aspect of the invention pertains to methods of making an antibody that binds human interleukin-18 (IL-18). The invention provides a method comprising
- 35 exposing an antibody repertoire to an antigen comprising an epitope of human IL-18 comprising amino acids PLFEDMTDSDCRDNA (SEQ ID NO: 1), VIRNLNDQVLFIDQ (SEQ ID NO: 33), or a portion of either; and selecting from the antibody repertoire an antibody that binds the epitope of human IL-18 comprising amino

acids PLFEDMTDSDCRDNA (SEQ ID NO: 1), VIRNLNDQVLFIDQ (SEQ ID NO: 33), or a portion of either.

In one embodiment, the antibody repertoire is an *in vivo* repertoire in an animal and the method comprises immunizing the animal with the antigen comprising the epitope of human IL-18 comprising amino acids PLFEDMTDSDCRDNA (SEQ ID NO: 1), VIRNLNDQVLFIDQ (SEQ ID NO: 33), the N- or C-terminal portion of human IL-18 (SEQ ID NOS: 70-71), or a portion of any of these epitopes. In another embodiment, the antibody repertoire is a recombinant antibody library and the method comprises screening the library with an antigen containing the epitope of human IL-18 having the amino acids PLFEDMTDSDCRDNA (SEQ ID NO: 1), VIRNLNDQVLFIDQ (SEQ ID NO: 33), peptides represented by SEQ ID NOS: 31-32 and 34-60, or a portion of any of the foregoing. Preferably, the library is a human antibody library.

In another aspect, the invention provides an isolated nucleic acid encoding an antibody of any of the above aspects, *e.g.*, a heavy and/or light chain variable region, or portion thereof. In related embodiments, the isolated nucleic acid encoding the anti-IL-18 antibody, or portion thereof, is in a recombinant expression vector, *e.g.*, for expression in a host cell.

Thus, in another aspect, the invention pertains to a method of using the foregoing host cell into which the recombinant expression vector has been introduced, for synthesizing an antibody that binds human IL-18, by culturing the host cell in a culture medium until an antibody that binds human IL-18 is synthesized by the cell.

Another aspect of the invention pertains to a method for inhibiting human IL-18 activity comprising contacting human IL-18 with the antibody, or antigen-binding portion thereof, of the invention such that human IL-18 activity is inhibited.

Yet another aspect of the invention pertains to a method for inhibiting human IL-18 activity in a human subject suffering from a disorder in which IL-18 activity is detrimental, comprising administering to the human subject the antibody, or antigen-binding portion thereof, of the invention such that human IL-18 activity in the human subject is inhibited. In one embodiment, the anti-IL-18 antibody may be administered, *e.g.*, before, concurrent, or after, an additional agent such as an anti-IL-12 antibody or antigen binding fragment thereof, methotrexate, anti-TNF antibody or antigen binding fragment thereof, corticosteroids, cyclosporin, rapamycin, FK506, or a non-steroidal anti-inflammatory agent.

Brief Description of the Drawings

Figure 1 shows the structural model of IL-18 (center) as compared to IL-1 β (left) and IL1RA (right).

5 *Figure 2* shows a structural model of IL-18 complexed with the IL-18 receptor, wherein the peptide epitope comprising amino acids PLFEDMTDSDCRDNA (SEQ ID NO: 1) of IL-18 is indicated in dark gray. This peptide epitope is bound by the anti-IL-18 antibody 2E1.

10 *Figure 3* shows a structural model of IL-18 complexed with the IL-18 receptor, wherein the peptide epitope comprising amino acids YFGKLESKLSVIRN (SEQ ID NO: 33) of IL-18 is indicated in dark gray. This peptide epitope is bound by the anti-IL-18 antibody LT28.

15 *Figure 4* shows a structural model of full length IL-18 complexed with the IL-18 receptor. The spherical light and dark gray epitopes represent the N and C terminal contact epitopes of IL-18 (respectively, SEQ ID NOS: 70 and 71).

20 *Figure 5* shows the potency of three different anti-IL-18 antibodies in neutralizing the biologic effects of IL-18 as a function of inhibition of IFN- γ induction in KG1 cells. The IC50 values for the antibodies 125H (boxes) and the 2E1 antibody as an IgG antibody (circles) or as a single chain antibody (triangles) are, respectively, 2.1E-10, 9.0E-10, and 3.3E-9.

Detailed Description of the Invention

25 This invention pertains to the selection of peptide epitopes that are capable of generating neutralizing antibodies to IL-18 mediated signal transduction, the preparation of antibodies to these epitopes and the use of such antibodies, including use to treat disorders involving IL-18. The strategy of selecting epitopes entails construction of an
30 homology model of the IL-18 protein and its corresponding receptor. A combination of visual inspection and computational evaluation is then used to select representative peptide segments for synthesis and antibody generation. Amino acid sequences shown herein use the standard one-letter abbreviation code.

35 Selection of IL-18 Epitopes

 The program Modeler (Sali, A. et al., *Evaluation of comparative protein modeling by MODELLER*. Proteins: Struct., Funct., Genet. (1995), 23(3), pp. 318-26. CODEN: PSFGEY; ISSN: 0887-3585) was used to generate homology models for both

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IL-18 and the IL-18 receptor. The X-ray crystal structures of IL-1 β (Priestle, J., et al. *The three-dimensional structure of human interleukin-1.beta. refined to 2.0 .ANG. resolution.* Prog. Clin. Biol. Res. (1990), 349 (Cytokines Lipocortins Inflammation Differ.), pp. 297-307) and IL-1RA (Schreuder, H. et al., *Refined crystal structure of the interleukin-1 receptor antagonist: presence of a disulfide link and a cis-proline.* Eur. J. Biochem. (1995), 227(3), pp. 838-47) are available and were used as reference coordinates for the model construction of IL-18. The IL-1 receptor structure (Vigers, G., et al. *Crystal structure of the type-1 interleukin-1-receptor complexed with interleukin-1.beta.* Nature (London) (1997), 386(6621), pp. 190-194) was used to model the IL-18 receptor.

The structural model building for IL-18 and the IL-18 receptor is described further below.

IL-18 Model Building

The overall sequence homology with these two proteins (*i.e.*, IL-1 β and IL-18) is low, however, there is compelling evidence that IL-18 is a member of the IL-1 family (see Dinarello, C.A. *IL-18: a TH1-inducing, proinflammatory cytokine and new member of the IL-1 family.* J. Allergy Clin. Immunol. (1999), 103(1, Pt. 1), pp. 11-24) and that the overall protein fold is very similar. Like IL-1 β , IL-18 is initially secreted in a pro form. Both pro-IL-1 β and pro-IL-18 are activated by IL-1 β -converting enzyme (ICE) (Fantuzzi, G. and Dinarello, C.A. *Interleukin-18 and interleukin-1 β : two cytokine substrates for ICE (caspase-1).* J. Clin. Immunol. (1999), 19(1), pp. 1-11). It is also known that the IL-1 receptor and the IL-18 receptor are similar (Dinarello, C.A. et al. *Overview of interleukin-18: more than an interferon- γ inducing factor.* J. Leukocyte Biol. (1998), 63(6), 658-664). IL-1 β is capable of binding to the IL-18 receptor. As a final argument, IL-1 β and IL-1RA display an identical fold, even though the overall sequence homology between these two proteins is on a par with the sequence homology with IL-18. The sequence alignment between the three proteins (*i.e.*, IL-18, IL-1 β and IL-1RA) was constructed manually with the program InsightII. This alignment can be seen in Table 1:

Table 1: Sequence alignment for IL-18 relative to IL-1 β and IL-1RA

5	IL-18:	24	YFGK-LESKLS-VIRNLNDQVLFIDQGNRPLFE--DMT-DSDCRD--NAP
	IL-1 β :		AP-VRS-LNCTLRDSQQKSLVMS-G--P-YELKALHLQGQ--D--MEQ
	IL-1RA:		SSKMQA-FR--IWDVNQKTFYLR-N--N--QLVAGYLQGP--NVNLEE
10	IL-18:	80	RTIFIISMY-KDSQPRG-MAVTISVKCEKISTLSC----ENK-IISFKEM
	IL-1 β :		QVVFSMS-FVQGEESNDKIPVALGLK-EKNLYLSCVLK-DDKPTLQLESV
	IL-1RA:		KI--DV---VP-IEPH---ALFLGIH-GGKMCLSCV-KSGDETRLQLEAV
15	IL-18:	123	NPPDNI-KDTKSDIIF-FQRSVPGHDNKMQFESSSYEGYFLACE-KERDL
	IL-1 β :		DPKNYP-KK-KMEKRFVFNK-I-EINNKLFEFSAQFPNWIYSTS-QAENM
	IL-1RA:		NITDLSNR-KQDKRFAFIR-S-DSGPTTSFESAACPGWFLCTAMEADQ-
20	IL-18:	170	FKLILKKED-ELGDRSIM-FTVQNE
	IL-1 β :		-PVFL--GG-TKGGQDITDFTMQFVSS
	IL-1RA:		-PVSL--TNMPDEGVMVTKFYFQED

(SEQ ID NO: 4)

(SEQ ID NO: 5)

(SEQ ID NO: 6)

25 The sequence homology between these sequences is listed in Table 2. The upper triangle is percent strict sequence identity and the lower triangle is percent conservative sequence homology. Only the portions of the total sequences reported in Table 1 are considered in Table 2. As was mentioned above, the overall homology is low but consistent across the family.

30 Table 2: Sequence homology between IL-1 family members

Molecule	IL-18	IL-1 β	IL-1RA	IL-1 Rec	IL-18 Rec
IL-18	-	20.0	21.8	-	-
IL-1b	53.5	-	27.5	-	-
IL-1RA	50.6	54.4	-	-	26.1
IL-1 Rec	-	-	-	-	50.5
IL-18 Rec	-	-	-	50.5	-

35 The resulting IL-18 structure is pictured in Figure 1 along with IL-1 β and IL-1RA. The overall quality as assessed by the program What_Check (Hooft, R.W. et al., *Errors in Protein Structures*. Nature (1996) 381, pp. 272) is reasonable, but a bit low (see Table 3 below).

Table 3: Structural Z-scores from What_Check (Positive is better than average)

	IL-1 β	IL-1RA	IL-1 Rec	IL-18 (model)	IL-18 Rec (model)
Packing Quality	-1.6	-2.3	-3.6	-5.5	-5.7
Ramachandran Plot	-2.0	-1.3	-2.9	-3.3	-2.3
Rotamer Normality	-1.6	-0.8	-1.5	-0.6	-0.6
Backbone Conformation	-1.7	+0.5	-1.6	-5.6	-2.7

5 However, the assessment of the reference structures by What_Check is also low, suggesting that this protein fold is poorly represented in the database of reference structures. Undoubtedly though, the low sequence homology contributed to a less than perfect final structure in spite of our confidence in the overall protein fold. However, for the purpose of choosing epitopes for antibody generation, this structure is considered to be sufficient.

IL-18 Receptor Model Building

15 The structure of the IL-18 receptor was also generated using the program Modeler. The reference coordinates were from the IL-1 receptor. As in the case of the cytokines associated with these receptors, the overall sequence identity is low, but sufficient to generate an alignment. The sequence homology figures are included in Table 2 above. The alignment was generated manually using the program InsightII and is presented in Table 4.

20 Table 4. Sequence alignment for the IL-18 receptor relative to the IL-1 Receptor

	22	IL-18 Rec: CTSRPHITVVEGEPFYLKHCSCSLAHEIETTTKSWYKSSGSQEHVELNPR
		IL-1 Rec: CKEREEKIILVSSANEIDVRPCPLNPNEHKGTITWYKDD-SKTPVSTEQA
	72	
		IL-18 Rec: SSSRIALHDCVLEFWPVELNDTGSYFFQMKNYTQKWKLNVIRRNKHS---
25		IL-1 Rec: S--RIHQHKEKLWFVPAKVEDSGHYVCVVRNSSYCLRIKISAKFVENEPN
	119	
		IL-18 Rec: -CFTERQVTSKIVEVKKFFQITCENSYYQTLVNST----SLYKNCKKLLL
		IL-1 Rec: LCYNAQAIFKQKLPVAGDGLVCPYMEFFKNENNELPKLQWYKDCCKPLLL
	163	
		IL-18 Rec: EN-----NKNPTIKKNAEFEDQGYYS CVHFLHNGKLFNITKTFNITIVE
		IL-1 Rec: DNIHFGVKDRILIVMNVAEKHRGNYTCHASYTLGKQYPITRVIEFITLE
	209	
30		IL-18 Rec: DRSNIVPVLLGPKLNHVAVELGKNVRLNCSALLNEEDVIYWMF-GEE-NG
		IL-1 Rec: ENKPTRPVIVSPANETMEVDLGSQIQLICNVGTGQLSDIAYWKWNGSVIDE
	257	
		IL-18 Rec: SDPNIHEE-KEMRIMTPEGKWHASKVLRIENIGESNLNVLYNCTVASTGG
		IL-1 Rec: DDPVLGEDYYSVENPANKRRSTLITVLNISEIESRFYKHPFTCFAKNTHG
	306	
		IL-18 Rec: TDTKSFILVRKAD (SEQ ID NO: 7)
		IL-1 Rec: IDAAYIQLIYPVT (SEQ ID NO: 8)

5 The overall quality of the structure of the IL-18 receptor as determined using the
Modeler program is reasonable but again scores somewhat low according to
What_Check (see Table 3 above). The confidence that can be placed on the overall fold
comes primarily from the fact that IL-1 β binds to both the IL-1 and IL-18 receptor. The
low sequence homology certainly contributes to the quality of the final structure,
10 however, as in the case of the associated cytokines above, this current structure is
considered to be sufficient. As an additional exercise, the IL-18 peptide epitope bound
by LT28 (SEQ ID NO: 33) was modeled when complexed with the IL-18 receptor
(Figure 3). As a final exercise, a model of the IL-18/IL-18 receptor complex was
generated based on the IL-1 β /IL-1 receptor structure (Figure 4). This structure was
15 generated by superimposing the cytokine structures and the receptor structures. No
attempt was made to energy minimize the final structure.

Peptide Epitope Selection

20 The primary purpose of generating structural models was to be able to select
suitable peptides based primarily on a visual score. Solvent exposed sections of the
proteins and portions of the proteins which were both hydrophilic and buried in the
receptor/cytokine complex were considered. A final element considered was based on a
selectivity criterion. The selected peptide epitope should be different in sequence from
similar portions of other members of the family. Based on this criteria, a peptide from
25 IL-18 was selected. In addition, a comprehensive overlapping panel of peptides (SEQ
ID NOS: 31-60) representative of full length human IL-18 (SEQ ID NO: 61) was also
made and the sequence of all of these IL-18 related peptides is shown in Table 5, below.

Table 5. Selected Peptides Representative of IL-18

30

Peptide Sequence	SEQ ID NO:
PLFEDMTDSDCRDNA	(SEQ ID NO: 1)
CPLFEDMTDSDCRDNA	(SEQ ID NO: 2)
PLFEDMTDSDCR	(SEQ ID NO: 3)
YFGKLESKLSVIRN	(SEQ ID NO: 31)
ESKLSVIRNLNDQV	(SEQ ID NO: 32)
VIRNLNDQVLFIDQ (LT28 binding epitope)	(SEQ ID NO: 33)
NDQVLFIDQGNRPL	(SEQ ID NO: 34)
FIDQGNRPLFEDMT	(SEQ ID NO: 35)

NRPLFEDMTDSDCR (2E1 binding epitope)	(SEQ ID NO: 36)
EDMTDSDCRDNAPR	(SEQ ID NO: 37)
SDCRDNAPRTIFII	(SEQ ID NO: 38)
NAPRTIFIISMYKD	(SEQ ID NO: 39)
IFIISMYKDSQPRG	(SEQ ID NO: 40)
MYKDSQPRGMAVTI	(SEQ ID NO: 41)
QPRGMAVTISVKCE	(SEQ ID NO: 42)
AVTISVKCEKISTL	(SEQ ID NO: 43)
VKCEKISTLSCENK	(SEQ ID NO: 44)
ISTLSCENKIISFK	(SEQ ID NO: 45)
CENKIISFKEMNPP	(SEQ ID NO: 46)
ISFKEMNPPDNIKD	(SEQ ID NO: 47)
MNPPDNIKDTKSDI	(SEQ ID NO: 48)
NIKDTKSDI IFFQR	(SEQ ID NO: 49)
KSDI IFFQRSVPGH	(SEQ ID NO: 50)
FFQRSVPGHDNKMQ	(SEQ ID NO: 51)
VPGHDNKMQFESS	(SEQ ID NO: 52)
NKMQFESSSYEGYF	(SEQ ID NO: 53)
ESSSYEGYFLACEK	(SEQ ID NO: 54)
EGYFLACEKERDLF	(SEQ ID NO: 55)
ACEKERDLFKLILK	(SEQ ID NO: 56)
RDLFKLILKKEDEL	(SEQ ID NO: 57)
LILKKEDELGDRSI	(SEQ ID NO: 58)
EDELGDRSIMFTVQ	(SEQ ID NO: 59)
DRSIMFTVQNE	(SEQ ID NO: 60)
YFGKLESKLS VIRNLNDQVLFIDQGNRPLFEDMTDSDC RD NAPRTIFIISMYKDSQPRGMAVTISVKCEKISTLSC ENKIISFKEMNPPDNIKDTKSDI IFFQRSVPGHDNKMQ FESSSYEGYFLACEKERDLFKLILKKEDELGDRSIMFT VQNE	(SEQ ID NO: 61) LT28 and 2E1 epitopes indicated in bold

- The N-terminal cysteine of the IL-18 peptide represented by SEQ ID NO: 2 is not part of the native IL-18 sequence, but was added as a conjugation site. Accordingly, within the native IL-18 amino acid sequence, the region corresponding to the selected epitope comprises amino acid residues having the amino acid sequence
- 5 PLFEDMTDSDCRDNA (SEQ ID NO: 1).

A schematic model of the IL-18 peptide (SEQ ID NO: 1) complexed with the IL-18 receptor is shown in Figure 2, with this peptide epitope indicated in dark gray.

Subsequent antigenicity calculations were performed on IL-18 peptide sequences, with the result that this peptide scored particularly highly. This peptide was synthesized and used as an epitope to generate antibodies in a rabbit host. Molecular modeling data obtained using the IL-18 peptide PLFEDMTDSDCRDNA (SEQ ID NO: 1) or YFGKLESKLSVIRN (SEQ ID NO: 31) as compared to a cognate receptor, *i.e.*, the IL-18 receptor, as depicted in Figures 2 and 3, provides an indication as to what residues a neutralizing antibody or compound may interact with.

An alternative method for peptide epitope selection can be accomplished in the absence of any molecular modeling by screening a panel of representative peptides using immunoselection. In one approach, overlapping peptides representative of the entire protein sequence can be used. In a more limited approach, only certain epitopes are represented in the panel of peptides. In a combined approach, molecular modeling can be used to identify epitopes likely to be important. The identified epitope(s) sequence can then be used to construct a panel of peptides (*e.g.*, overlapping peptides) that are representative of the identified epitope(s). Methods for manufacturing desired peptide sequences can be carried out using standard techniques known in the art.

Once the binding peptide or peptides (*e.g.*, panel of overlapping peptides) has been selected, an immunoscreen for a cognate receptor can be performed. Alternatively, an immunoscreen can be performed with a selected cognate receptor such that a peptide having a certain binding affinity can be identified. Any number of immunoscreens can be employed such that, either a desired receptor or desired peptide can be identified as a candidate binding molecule for further study. Such "bait" and "prey" techniques for analyzing protein-protein interactions, for identifying candidate binding molecules, and/or for scoring binding affinities are described in the art. One preferred technique utilizes phage display as described herein.

Anti-IL-18 Antibodies

The invention provides antibodies, as well as antibody portions thereof, that bind IL-18. Preferably, the antibodies, or portions thereof, are isolated antibodies. Preferably, the antibodies, or portions thereof, are neutralizing antibodies.

The term "antibody", as used herein, is intended to refer to immunoglobulin molecules comprised of four polypeptide chains, two heavy (H) chains and two light (L) chains inter-connected by disulfide bonds. Each heavy chain is comprised of a heavy chain variable region (abbreviated herein as HCVR or VH) and a heavy chain constant region. The heavy chain constant region is comprised of three domains, CH1, CH2 and CH3. Each light chain is comprised of a light chain variable region (abbreviated herein as LCVR or VL) and a light chain constant region. The light chain constant region is comprised of one domain, CL. The VH and VL regions can be further subdivided into

regions of hypervariability, termed complementarity determining regions (CDR), interspersed with regions that are more conserved, termed framework regions (FR). Each VH and VL is composed of three CDRs and four FRs, arranged from amino-terminus to carboxy-terminus in the following order: FR1, CDR1, FR2, CDR2, FR3,

5 CDR3, FR4.

The term "antigen-binding portion" of an antibody (or simply "antibody portion"), as used herein, refers to one or more fragments of an antibody that retain the ability to specifically bind to an antigen (*e.g.*, hIL-18). It has been shown that the antigen-binding function of an antibody can be performed by fragments of a full-length

10 antibody. Examples of binding fragments encompassed within the term "antigen-binding portion" of an antibody include (i) a Fab fragment, a monovalent fragment consisting of the VL, VH, CL and CH1 domains; (ii) a F(ab')₂ fragment, a bivalent fragment comprising two Fab fragments linked by a disulfide bridge at the hinge region; (iii) a Fd fragment consisting of the VH and CH1 domains; (iv) a Fv fragment consisting

15 of the VL and VH domains of a single arm of an antibody, (v) a dAb fragment (Ward *et al.*, (1989) *Nature* 341:544-546), which consists of a VH domain; and (vi) an isolated complementarity determining region (CDR). Furthermore, although the two domains of the Fv fragment, VL and VH, are coded for by separate genes, they can be joined, using recombinant methods, by a synthetic linker that enables them to be made as a single

20 protein chain in which the VL and VH regions pair to form monovalent molecules (known as single chain Fv (scFv); see *e.g.*, Bird *et al.* (1988) *Science* 242:423-426; and Huston *et al.* (1988) *Proc. Natl. Acad. Sci. USA* 85:5879-5883). Such single chain antibodies are also intended to be encompassed within the term "antigen-binding portion" of an antibody. Other forms of single chain antibodies, such as diabodies are

25 also encompassed. Diabodies are bivalent, bispecific antibodies in which VH and VL domains are expressed on a single polypeptide chain, but using a linker that is too short to allow for pairing between the two domains on the same chain, thereby forcing the domains to pair with complementary domains of another chain and creating two antigen binding sites (see *e.g.*, Holliger, P., *et al.* (1993) *Proc. Natl. Acad. Sci. USA* 90:6444-

30 6448; Poljak, R.J., *et al.* (1994) *Structure* 2:1121-1123).

Still further, an antibody or antigen-binding portion thereof may be part of a larger immunoadhesion molecules, formed by covalent or noncovalent association of the antibody or antibody portion with one or more other proteins or peptides. Examples of such immunoadhesion molecules include use of the streptavidin core region to make a

35 tetrameric scFv molecule (Kipriyanov, S.M., *et al.* (1995) *Human Antibodies and Hybridomas* 6:93-101) and use of a cysteine residue, a marker peptide and a C-terminal polyhistidine tag to make bivalent and biotinylated scFv molecules (Kipriyanov, S.M., *et al.* (1994) *Mol. Immunol.* 31:1047-1058). Antibody portions, such as Fab and F(ab')₂

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fragments, can be prepared from whole antibodies using conventional techniques, such as papain or pepsin digestion, respectively, of whole antibodies. Moreover, antibodies, antibody portions and immunoadhesion molecules can be obtained using standard recombinant DNA techniques, as described herein.

5 An "isolated antibody", as used herein, is intended to refer to an antibody that is substantially free of other antibodies having different antigenic specificities (*e.g.*, an isolated antibody that specifically binds hIL-18 is substantially free of antibodies that specifically bind antigens other than hIL-18). An isolated antibody that specifically binds hIL-18 may, however, have cross-reactivity to other antigens, such as IL-18
10 molecules from other species. Moreover, an isolated antibody may be substantially free of other cellular material and/or chemicals. Further, an isolated antibody, *e.g.*, an isolated human antibody, can be a chimeric antibody wherein, *e.g.*, variable regions, CDR domains, or isotypes derived from a different human source are grafted to the parent human antibody.

15 A "compound" as used herein, refers to binding molecules such as antibodies, *e.g.*, polyclonal antibodies, monoclonal antibodies, binding fragments thereof (*e.g.*, Fab fragments), single chain antibodies (*e.g.*, scFv), peptides or peptide mimetics, as well as non-peptide based molecules, such as small molecules having ligand binding activity.

A "neutralizing antibody", as used herein (or an "antibody that neutralized hIL-
20 18 activity"), is intended to refer to an antibody whose binding to hIL-18 results in inhibition of the biological activity of hIL-18. This inhibition of the biological activity of hIL-18 can be assessed by measuring one or more indicators of hIL-18 biological activity. These indicators of hIL-18 biological activity can be assessed by one or more of several standard *in vitro* or *in vivo* assays known in the art.

25 The term "surface plasmon resonance", as used herein, refers to an optical phenomenon that allows for the analysis of real-time biospecific interactions by detection of alterations in protein concentrations within a biosensor matrix, for example using the BIAcore system (Pharmacia Biosensor AB, Uppsala, Sweden and Piscataway, NJ). For further descriptions, see Jönsson, U., *et al.* (1993) *Ann. Biol. Clin.* 51:19-26;
30 Jönsson, U., *et al.* (1991) *Biotechniques* 11:620-627; Johnsson, B., *et al.* (1995) *J. Mol. Recognit.* 8:125-131; and Johnsson, B., *et al.* (1991) *Anal. Biochem.* 198:268-277.

The term " K_{off} ", as used herein, is intended to refer to the off rate constant for dissociation of an antibody from the antibody/antigen complex.

The term " K_d ", as used herein, is intended to refer to the dissociation constant of
35 a particular antibody-antigen interaction.

In one aspect, the invention pertains to an isolated antibody, or an antigen-binding portion thereof, that binds an epitope of human IL-18 comprising amino acids PLFEDMTDSDCRDNA (SEQ ID NO: 1) or VIRNLNDQVLFIDQ (SEQ ID NO: 33),

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or a portion of any of these epitopes. Preferably, the antibody is a neutralizing antibody. Preferably, the antibody is a human antibody. In various embodiments, the antibody is a recombinant antibody or a monoclonal antibody.

In other embodiments, the isolated antibody, or antigen-binding portion thereof, binds to an epitope of human IL-18 comprising amino acids PLFEDMTDSDCRDNA (SEQ ID NO: 1), wherein the antibody, or antigen-binding portion thereof, dissociates from human IL-18 with a k_{off} rate constant of 0.1s^{-1} or less, as determined by surface plasmon resonance, or which inhibits human IL-18 activity with an IC_{50} of $1 \times 10^{-6}\text{M}$ or less. Alternatively, the antibody, or an antigen-binding portion thereof, may dissociate from human IL-18 with a k_{off} rate constant of $1 \times 10^{-2}\text{s}^{-1}$ or less, as determined by surface plasmon resonance, or may inhibit human IL-18 activity with an IC_{50} of $1 \times 10^{-7}\text{M}$ or less. Alternatively, the antibody, or an antigen-binding portion thereof, may dissociate from human IL-18 with a k_{off} rate constant of $1 \times 10^{-3}\text{s}^{-1}$ or less, as determined by surface plasmon resonance, or may inhibit human IL-18 activity with an IC_{50} of $1 \times 10^{-8}\text{M}$ or less. Alternatively, the antibody, or an antigen-binding portion thereof, may dissociate from human IL-18 with a k_{off} rate constant of $1 \times 10^{-4}\text{s}^{-1}$ or less, as determined by surface plasmon resonance, or may inhibit human IL-18 activity with an IC_{50} of $1 \times 10^{-9}\text{M}$ or less. Alternatively, the antibody, or an antigen-binding portion thereof, may dissociate from human IL-18 with a k_{off} rate constant of $1 \times 10^{-5}\text{s}^{-1}$ or less, as determined by surface plasmon resonance, or may inhibit human IL-18 activity with an IC_{50} of $1 \times 10^{-10}\text{M}$ or less. Alternatively, the antibody, or an antigen-binding portion thereof, may dissociate from human IL-18 with a k_{off} rate constant of $1 \times 10^{-6}\text{s}^{-1}$ or less, as determined by surface plasmon resonance, or may inhibit human IL-18 activity with an IC_{50} of $1 \times 10^{-11}\text{M}$ or less.

Affinity Maturation of Identified Anti-IL-18 Antibodies

The invention also provides for the further modification of an antibody identified as binding to an IL-18 epitope. The modification of the identified anti-IL-18 antibody is to improve binding and/or neutralization activity.

Therapeutic Compositions and Methods for Administering

The invention also provides pharmaceutical compositions comprising an antibody, or antigen-binding portion thereof, of the invention and a pharmaceutically acceptable carrier. In one embodiment, the pharmaceutical composition further comprises at least one additional therapeutic agent for treating a disorder in which IL-18 activity is detrimental.

The antibodies and antibody-portions of the invention can be incorporated into pharmaceutical compositions suitable for administration to a subject. Typically, the

pharmaceutical composition comprises an antibody or antibody portion of the invention and a pharmaceutically acceptable carrier. As used herein, "pharmaceutically acceptable carrier" includes any and all solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents, and the like that are physiologically compatible. Examples of pharmaceutically acceptable carriers include one or more of water, saline, phosphate buffered saline, dextrose, glycerol, ethanol and the like, as well as combinations thereof. In many cases, it will be preferable to include isotonic agents, for example, sugars, polyalcohols such as mannitol, sorbitol, or sodium chloride in the composition. Pharmaceutically acceptable carriers may further comprise minor amounts of auxiliary substances such as wetting or emulsifying agents, preservatives or buffers, which enhance the shelf life or effectiveness of the antibody or antibody portion.

The antibodies and antibody-portions of the invention can be incorporated into a pharmaceutical composition suitable for parenteral administration. Preferably, the antibody or antibody-portions will be prepared as an injectable solution containing 0.1-250 mg/ml antibody. The injectable solution can be composed of either a liquid or lyophilized dosage form in a flint or amber vial, ampule or pre-filled syringe. The buffer can be L-histidine (1-50 mM), optimally 5-10 mM, at pH 5.0 to 7.0 (optimally pH 6.0). Other suitable buffers include but are not limited to, sodium succinate, sodium citrate, sodium phosphate or potassium phosphate. Sodium chloride can be used to modify the toxicity of the solution at a concentration of 0-300 mM (optimally 150 mM for a liquid dosage form). Cryoprotectants can be included for a lyophilized dosage form, principally 0-10% sucrose (optimally 0.5-1.0%). Other suitable cryoprotectants include trehalose and lactose. Bulking agents can be included for a lyophilized dosage form, principally 1-10% mannitol (optimally 2-4%). Stabilizers can be used in both liquid and lyophilized dosage forms, principally 1-50 mM L-Methionine (optimally 5-10 mM). Other suitable bulking agents include glycine, arginine, can be included as 0-0.05% polysorbate-80 (optimally 0.005-0.01%). Additional surfactants include but are not limited to polysorbate 20 and BRIJ surfactants.

The compositions of this invention may be in a variety of forms. These include, for example, liquid, semi-solid and solid dosage forms, such as liquid solutions (e.g., injectable and infusible solutions), dispersions or suspensions, tablets, pills, powders, liposomes and suppositories. The preferred form depends on the intended mode of administration and therapeutic application. Typical preferred compositions are in the form of injectable or infusible solutions, such as compositions similar to those used for passive immunization of humans with other antibodies. The preferred mode of administration is parenteral (e.g., intravenous, subcutaneous, intraperitoneal, intramuscular). In a preferred embodiment, the antibody is administered by intravenous

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infusion or injection. In another preferred embodiment, the antibody is administered by intramuscular or subcutaneous injection.

Therapeutic compositions typically must be sterile and stable under the conditions of manufacture and storage. The composition can be formulated as a solution, microemulsion, dispersion, liposome, or other ordered structure suitable to high drug concentration. Sterile injectable solutions can be prepared by incorporating the active compound (*i.e.*, antibody or antibody portion) in the required amount in an appropriate solvent with one or a combination of ingredients enumerated above, as required, followed by filtered sterilization. Generally, dispersions are prepared by incorporating the active compound into a sterile vehicle that contains a basic dispersion medium and the required other ingredients from those enumerated above. In the case of sterile, lyophilized powders for the preparation of sterile injectable solutions, the preferred methods of preparation are vacuum drying and spray-drying that yields a powder of the active ingredient plus any additional desired ingredient from a previously sterile-filtered solution thereof. The proper fluidity of a solution can be maintained, for example, by the use of a coating such as lecithin, by the maintenance of the required particle size in the case of dispersion and by the use of surfactants. Prolonged absorption of injectable compositions can be brought about by including in the composition an agent that delays absorption, for example, monostearate salts and gelatin.

The antibodies and antibody-portions of the present invention can be administered by a variety of methods known in the art, although for many therapeutic applications, the preferred route/mode of administration is subcutaneous injection, intravenous injection or infusion. As will be appreciated by the skilled artisan, the route and/or mode of administration will vary depending upon the desired results. In certain embodiments, the active compound may be prepared with a carrier that will protect the compound against rapid release, such as a controlled release formulation, including implants, transdermal patches, and microencapsulated delivery systems. Biodegradable, biocompatible polymers can be used, such as ethylene vinyl acetate, polyanhydrides, polyglycolic acid, collagen, polyorthoesters, and polylactic acid. Many methods for the preparation of such formulations are patented or generally known to those skilled in the art. See, *e.g.*, *Sustained and Controlled Release Drug Delivery Systems*, J.R. Robinson, ed., Marcel Dekker, Inc., New York, 1978.

In certain embodiments, an antibody or antibody portion of the invention may be orally administered, for example, with an inert diluent or an assimilable edible carrier. The compound (and other ingredients, if desired) may also be enclosed in a hard or soft shell gelatin capsule, compressed into tablets, or incorporated directly into the subject's diet. For oral therapeutic administration, the compounds may be incorporated with excipients and used in the form of ingestible tablets, buccal tablets, troches, capsules,

elixirs, suspensions, syrups, wafers, and the like. To administer a compound of the invention by other than parenteral administration, it may be necessary to coat the compound with, or co-administer the compound with, a material to prevent its inactivation.

- 5 Supplementary active compounds can also be incorporated into the compositions. In certain embodiments, an antibody or antibody portion of the invention is coformulated with and/or coadministered with one or more additional therapeutic agents that are useful for treating disorders in which IL-18 activity is detrimental. For example, an anti-hIL-18 antibody or antibody portion of the invention may be
- 10 coformulated and/or coadministered with one or more additional antibodies that bind other targets (*e.g.*, antibodies that bind other cytokines or that bind cell surface molecules). Furthermore, one or more antibodies of the invention may be used in combination with two or more of the foregoing therapeutic agents. Such combination therapies may advantageously utilize lower dosages of the administered therapeutic
- 15 agents, thus avoiding possible toxicities or complications associated with the various monotherapies.

Therapeutic Uses

- 20 Interleukin 18 plays a critical role in the pathology associated with a variety of diseases involving immune and inflammatory elements. These diseases include, but are not limited to, rheumatoid arthritis, osteoarthritis, juvenile chronic arthritis, Lyme arthritis, psoriatic arthritis, reactive arthritis, spondyloarthropathy, systemic lupus erythematosus, Crohn's disease, ulcerative colitis, inflammatory bowel disease, insulin dependent diabetes mellitus, thyroiditis, asthma, allergic diseases, psoriasis, dermatitis
- 25 scleroderma, graft versus host disease, organ transplant rejection, acute or chronic immune disease associated with organ transplantation, sarcoidosis, atherosclerosis, disseminated intravascular coagulation, Kawasaki's disease, Grave's disease, nephrotic syndrome, chronic fatigue syndrome, Wegener's granulomatosis, Henoch-Schoenlein purpura, microscopic vasculitis of the kidneys, chronic active hepatitis, uveitis, septic
- 30 shock, toxic shock syndrome, sepsis syndrome, cachexia, infectious diseases, parasitic diseases, acquired immunodeficiency syndrome, acute transverse myelitis, Huntington's chorea, Parkinson's disease, Alzheimer's disease, stroke, primary biliary cirrhosis, hemolytic anemia, malignancies, heart failure, myocardial infarction, Addison's disease, sporadic, polyglandular deficiency type I and polyglandular deficiency type II, Schmidt's
- 35 syndrome, adult (acute) respiratory distress syndrome, alopecia, alopecia areata, seronegative arthropathy, arthropathy, Reiter's disease, psoriatic arthropathy, ulcerative colitic arthropathy, enteropathic synovitis, chlamydia, yersinia and salmonella associated arthropathy, spondyloarthropathy, atheromatous disease/arteriosclerosis,

- atopic allergy, autoimmune bullous disease, pemphigus vulgaris, pemphigus foliaceus, pemphigoid, linear IgA disease, autoimmune haemolytic anaemia, Coombs positive haemolytic anaemia, acquired pernicious anaemia, juvenile pernicious anaemia, myalgic encephalitis/Royal Free Disease, chronic mucocutaneous candidiasis, giant cell arteritis,
- 5 primary sclerosing hepatitis, cryptogenic autoimmune hepatitis, Acquired Immunodeficiency Disease Syndrome, Acquired Immunodeficiency Related Diseases, Hepatitis C, common varied immunodeficiency (common variable hypogammaglobulinaemia), dilated cardiomyopathy, female infertility, ovarian failure, premature ovarian failure, fibrotic lung disease, cryptogenic fibrosing alveolitis, post-
 - 10 inflammatory interstitial lung disease, interstitial pneumonitis, connective tissue disease associated interstitial lung disease, mixed connective tissue disease associated lung disease, systemic sclerosis associated interstitial lung disease, rheumatoid arthritis associated interstitial lung disease, systemic lupus erythematosus associated lung disease, dermatomyositis/polymyositis associated lung disease, Sjögren's disease
 - 15 associated lung disease, ankylosing spondylitis associated lung disease, vasculitic diffuse lung disease, haemosiderosis associated lung disease, drug-induced interstitial lung disease, radiation fibrosis, bronchiolitis obliterans, chronic eosinophilic pneumonia, lymphocytic infiltrative lung disease, postinfectious interstitial lung disease, gouty arthritis, autoimmune hepatitis, type-1 autoimmune hepatitis (classical autoimmune or
 - 20 lupoid hepatitis), type-2 autoimmune hepatitis (anti-LKM antibody hepatitis), autoimmune mediated hypoglycaemia, type B insulin resistance with acanthosis nigricans, hypoparathyroidism, acute immune disease associated with organ transplantation, chronic immune disease associated with organ transplantation, osteoarthritis, primary sclerosing cholangitis, psoriasis type 1, psoriasis type 2,
 - 25 idiopathic leucopaenia, autoimmune neutropaenia, renal disease NOS, glomerulonephritides, microscopic vasculitis of the kidneys, Lyme disease, discoid lupus erythematosus, male infertility idiopathic or NOS, sperm autoimmunity, multiple sclerosis (all subtypes), sympathetic ophthalmia, pulmonary hypertension secondary to connective tissue disease, Goodpasture's syndrome, pulmonary manifestation of
 - 30 polyarteritis nodosa, acute rheumatic fever, rheumatoid spondylitis, Still's disease, systemic sclerosis, Sjögren's syndrome, Takayasu's disease/arteritis, autoimmune thrombocytopaenia, idiopathic thrombocytopaenia, autoimmune thyroid disease, hyperthyroidism, goitrous autoimmune hypothyroidism (Hashimoto's disease), atrophic autoimmune hypothyroidism, primary myxoedema, phacogenic uveitis, primary
 - 35 vasculitis and vitiligo. The human antibodies, and antibody portions of the invention can be used to treat humans suffering from autoimmune diseases, in particular those associated with inflammation, including, rheumatoid spondylitis, allergy, autoimmune diabetes, autoimmune uveitis, acute liver disease, chronic liver diseases, allergy and

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asthma, mental disorders (*e.g.*, depression and schizophrenia), and Th2 Type and Th1 Type mediated diseases.

Preferably, the antibodies of the invention or antigen-binding portions thereof, are used to treat rheumatoid arthritis, Crohn's disease, multiple sclerosis, insulin
5 dependent diabetes, mellitus, and psoriasis.

An antibody, or antibody portion, of the invention also can be administered with one or more additional therapeutic agents useful in the treatment of autoimmune and inflammatory diseases.

Antibodies of the invention, or antigen binding portions thereof can be used
10 alone or in combination to treat such diseases. It should be understood that the antibodies of the invention or antigen binding portion thereof can be used alone or in combination with an additional agent, *e.g.*, a therapeutic agent, said additional agent being selected by the skilled artisan for its intended purpose. For example, the additional agent can be a therapeutic agent art-recognized as being useful to treat the
15 disease or condition being treated by the antibody of the present invention. The additional agent also can be an agent which imparts a beneficial attribute to the therapeutic composition *e.g.*, an agent which effects the viscosity of the composition.

It should further be understood that the combinations which are to be included within this invention are those combinations useful for their intended purpose. The
20 agents set forth below are illustrative for purposes and not intended to be limited. The combinations which are part of this invention can be the antibodies of the present invention and at least one additional agent selected from the lists below. The combination can also include more than one additional agent, *e.g.*, two or three additional agents if the combination is such that the formed composition can perform its
25 intended function.

Preferred combinations are non-steroidal anti-inflammatory drug(s) also referred to as NSAIDS which include drugs like ibuprofen. Other preferred combinations are corticosteroids including prednisolone; the well known side-effects of steroid use can be
30 reduced or even eliminated by tapering the steroid dose required when treating patients in combination with the anti-IL-18 antibodies of this invention. Non-limiting examples of therapeutic agents for rheumatoid arthritis with which an antibody, or antibody portion, of the invention can be combined include the following: cytokine suppressive anti-inflammatory drug(s) (CSAIDs); antibodies to or antagonists of other human cytokines or growth factors, for example, TNF, LT, IL-1, IL-2, IL-6, IL-7, IL-8, IL-12,
35 IL-15, IL-16, EMAP-II, GM-CSF, FGF, and PDGF. Antibodies of the invention, or antigen binding portions thereof, can be combined with antibodies to cell surface molecules such as CD2, CD3, CD4, CD8, CD25, CD28, CD30, CD40, CD45, CD69, CD80 (B7.1), CD86 (B7.2), CD90, or their ligands including CD154 (gp39 or CD40L).

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Preferred combinations of therapeutic agents may interfere at different points in the autoimmune and subsequent inflammatory cascade; preferred examples include TNF antagonists like chimeric, humanized or human TNF antibodies, D2E7, (PCT Publication No. WO 97/29131), CA2 (Remicade™), CDP 571, and soluble p55 or p75 TNF receptors, derivatives, thereof, (p75TNFR1gG (Enbrel™) or p55TNFR1gG (Lenercept)), and also TNF α converting enzyme (TACE) inhibitors; similarly IL-1 inhibitors (Interleukin-1-converting enzyme inhibitors, IL-1RA *etc.*) may be effective for the same reason. Other preferred combinations include Interleukin 11. Yet another preferred combination are other key players of the autoimmune response which may act parallel to, dependent on or in concert with IL-18 function; especially preferred are IL-12 antagonists including IL-12 antibodies or soluble IL-12 receptors, or IL-12 binding proteins. It has been shown that IL-12 and IL-18 have overlapping but distinct functions and a combination of antagonists to both may be most effective. Yet another preferred combination are non-depleting anti-CD4 inhibitors. Yet other preferred combinations include antagonists of the co-stimulatory pathway CD80 (B7.1) or CD86 (B7.2) including antibodies, soluble receptors or antagonistic ligands.

The antibodies of the invention, or antigen binding portions thereof, may also be combined with agents, such as methotrexate, 6-MP, azathioprine sulphasalazine, mesalazine, olsalazine chloroquine/hydroxychloroquine, pencillamine, aurothiomalate (intramuscular and oral), azathioprine, cochlincine, corticosteroids (oral, inhaled and local injection), beta-2 adrenoreceptor agonists (salbutamol, terbutaline, salmeteral), xanthines (theophylline, aminophylline), cromoglycate, nedocromil, ketotifen, ipratropium and oxitropium, cyclosporin, FK506, rapamycin, mycophenolate mofetil, leflunomide, NSAIDs, for example, ibuprofen, cox-2 inhibitors, cox-2 selective inhibitors (*e.g.*, rofecoxib (VIOXX™; Merck & Co., Inc.)) corticosteroids such as prednisolone, phosphodiesterase inhibitors, adenosine agonists, antithrombotic agents, complement inhibitors, adrenergic agents, agents which interfere with signalling by proinflammatory cytokines such as TNF α or IL-1 (*e.g.* IRAK, NIK, IKK, p38 or MAP kinase inhibitors), IL-1 β converting enzyme inhibitors, TNF α converting enzyme (TACE) inhibitors, T-cell signalling inhibitors such as kinase inhibitors, metalloproteinase inhibitors, sulfasalazine, azathioprine, 6-mercaptopurines, angiotensin converting enzyme inhibitors, soluble cytokine receptors and derivatives thereof (*e.g.* soluble p55 or p75 TNF receptors and the derivatives p75TNFR1gG (Enbrel™ and p55TNFR1gG (Lenercept)), sIL-1RI, sIL-1RII, sIL-6R) and antiinflammatory cytokines (*e.g.* IL-4, IL-10, IL-11, IL-13 and TGF β). Preferred combinations include methotrexate or leflunomide and in moderate or severe rheumatoid arthritis cases, cyclosporine.

Non-limiting examples of therapeutic agents for inflammatory bowel disease with which an antibody, or antibody portion, of the invention can be combined include

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Non-limiting examples of therapeutic agents for multiple sclerosis with which an antibody, or antibody portion, of the invention can be combined include the following:

- corticosteroids; prednisolone; methylprednisolone; azathioprine; cyclophosphamide; cyclosporine; methotrexate; 4-aminopyridine; tizanidine; interferon- β 1a (Avonex; Biogen); interferon- β 1b (Betaseron; Chiron/Berlex); Copolymer 1 (Cop-1; Copaxone; Teva Pharmaceutical Industries, Inc.); hyperbaric oxygen; intravenous immunoglobulin;
- 5 clabribine; antibodies to or antagonists of other human cytokines or growth factors, for example, TNF, LT, IL-1, IL-2, IL-6, IL-7, IL-8, IL-12, IL-15, IL-16, EMAP-II, GM-CSF, FGF, and PDGF. Antibodies of the invention, or antigen binding portions thereof, can be combined with antibodies to cell surface molecules such as CD2, CD3, CD4, CD8, CD25, CD28, CD30, CD40, CD45, CD69, CD80, CD86, CD90 or their ligands.
- 10 The antibodies of the invention, or antigen binding portions thereof, may also be combined with agents, such as methotrexate, cyclosporine, FK506, rapamycin, mycophenolate mofetil, leflunomide, NSAIDs, for example, ibuprofen, corticosteroids such as prednisolone, phosphodiesterase inhibitors, adenosine agonists, antithrombotic agents, complement inhibitors, adrenergic agents, agents which interfere with signalling
- 15 by proinflammatory cytokines such as TNF α or IL-1 (*e.g.* IRAK, NIK, IKK, p38 or MAP kinase inhibitors), IL-1 β converting enzyme inhibitors, TACE inhibitors, T-cell signalling inhibitors such as kinase inhibitors, metalloproteinase inhibitors, sulfasalazine, azathioprine, 6-mercaptopurines, angiotensin converting enzyme inhibitors, soluble cytokine receptors and derivatives thereof (*e.g.* soluble p55 or p75
- 20 TNF receptors, sIL-1RI, sIL-1RII, sIL-6R) and antiinflammatory cytokines (*e.g.* IL-4, IL-10, IL-13 and TGF β).

Preferred examples of therapeutic agents for multiple sclerosis in which the antibody or antigen binding portion thereof can be combined to include interferon- β , for example, IFN β 1a and IFN β 1b; copaxone, corticosteroids, IL-1 inhibitors, TNF

- 25 inhibitors, and antibodies to CD40 ligand and CD80.

The pharmaceutical compositions of the invention may include a "therapeutically effective amount" or a "prophylactically effective amount" of an antibody or antibody portion of the invention. A "therapeutically effective amount" refers to an amount effective, at dosages and for periods of time necessary, to achieve the desired therapeutic

30 result. A therapeutically effective amount of the antibody or antibody portion may vary according to factors such as the disease state, age, sex, and weight of the individual, and the ability of the antibody or antibody portion to elicit a desired response in the individual. A therapeutically effective amount is also one in which any toxic or detrimental effects of the antibody or antibody portion are outweighed by the

35 therapeutically beneficial effects. A "prophylactically effective amount" refers to an amount effective, at dosages and for periods of time necessary, to achieve the desired prophylactic result. Typically, since a prophylactic dose is used in subjects prior to or at

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an earlier stage of disease, the prophylactically effective amount will be less than the therapeutically effective amount.

Dosage regimens may be adjusted to provide the optimum desired response (e.g., a therapeutic or prophylactic response). For example, a single bolus may be administered, several divided doses may be administered over time or the dose may be proportionally reduced or increased as indicated by the exigencies of the therapeutic situation. It is especially advantageous to formulate parenteral compositions in dosage unit form for ease of administration and uniformity of dosage. Dosage unit form as used herein refers to physically discrete units suited as unitary dosages for the mammalian subjects to be treated; each unit containing a predetermined quantity of active compound calculated to produce the desired therapeutic effect in association with the required pharmaceutical carrier. The specification for the dosage unit forms of the invention are dictated by and directly dependent on (a) the unique characteristics of the active compound and the particular therapeutic or prophylactic effect to be achieved, and (b) the limitations inherent in the art of compounding such an active compound for the treatment of sensitivity in individuals.

An exemplary, non-limiting range for a therapeutically or prophylactically effective amount of an antibody or antibody portion of the invention is 0.1-20 mg/kg, more preferably 1-10 mg/kg. It is to be noted that dosage values may vary with the type and severity of the condition to be alleviated. It is to be further understood that for any particular subject, specific dosage regimens should be adjusted over time according to the individual need and the professional judgment of the person administering or supervising the administration of the compositions, and that dosage ranges set forth herein are exemplary only and are not intended to limit the scope or practice of the claimed composition.

Methods of Making Anti-IL-18 Antibodies

The anti-IL-18 antibodies of the invention are made using any one of a variety of techniques known in the art for preparing antibodies and using antigens comprising the IL-18 peptide epitope described in subsection I, i.e., an epitope of human IL-18 comprising amino acids PLFEDMTDSDCRDNA (SEQ ID NO: 1).

In general, the methods of the invention for making an antibody that binds human interleukin-18 (IL-18) involve:

exposing an antibody repertoire to an antigen comprising an epitope of human IL-18 comprising amino acids PLFEDMTDSDCRDNA (SEQ ID NO: 1), or portion thereof (e.g., SEQ ID NO: 3 or 33); and

selecting from the antibody repertoire an antibody that binds the epitope of human IL-18 comprising amino acids PLFEDMTDSDCRDNA (SEQ ID NO: 1), or portion thereof (*e.g.*, SEQ ID NO: 3 or 33).

- In one embodiment, the antibody repertoire is an *in vivo* repertoire in an animal and the method comprises immunizing the animal with the antigen comprising the epitope of human IL-18 comprising amino acids PLFEDMTDSDCRDNA (SEQ ID NO: 1). In another embodiment, the antibody repertoire is a recombinant antibody library and the method comprising screening the library with the antigen comprising the epitope of human IL-18 comprising amino acids PLFEDMTDSDCRDNA (SEQ ID NO: 1).
- 10 Preferably, the library is a human antibody library.

- Methods for immunizing an animal with an antigen to thereby raise specific antibodies to the antigen are well known in the art. An IL-18 antigen comprising an epitope of human IL-18 comprising amino acids PLFEDMTDSDCRDNA (SEQ ID NO: 1) can be administered to an animal to elicit polyclonal antibodies and specific antibodies that bind the epitope can be isolated by selecting from the polyclonal antibodies those antibodies that bind to the epitope (*e.g.*, by passing the polyclonal antisera over a column that comprises a peptide comprising amino acids PLFEDMTDSDCRDNA (SEQ ID NO: 1) of hIL-18). The antigen used to elicit the polyclonal antibodies can be intact (*i.e.*, full-length) hIL-18 or can be a portion of hIL-18 that includes the epitope of interest, *e.g.*, a synthetic peptide comprising amino acids PLFEDMTDSDCRDNA (SEQ ID NO: 1) of hIL-18. Furthermore, monoclonal antibodies to the epitope can be made from the aforementioned animals using standard hybridoma technology and selection for those hybridomas secreting an antibody that specifically binds the epitope of interest, *e.g.*, by screening the hybridomas with a peptide comprising amino acids PLFEDMTDSDCRDNA (SEQ ID NO: 1) of hIL-18 and selecting for antibodies that bind specifically to the peptide.
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- In vitro* methods also can be used to make the antibodies of the invention, wherein an antibody library is screened to identify an antibody having the desired binding specificity. Methods for such screening of recombinant antibody libraries are well known in the art and include methods described in, for example, Ladner *et al.* U.S. Patent No. 5,223,409; Kang *et al.* PCT Publication No. WO 92/18619; Dower *et al.* PCT Publication No. WO 91/17271; Winter *et al.* PCT Publication No. WO 92/20791; Markland *et al.* PCT Publication No. WO 92/15679; Breitling *et al.* PCT Publication No. WO 93/01288; McCafferty *et al.* PCT Publication No. WO 92/01047; Garrard *et al.* PCT Publication No. WO 92/09690; Fuchs *et al.* (1991) *Bio/Technology* 9:1370-1372; Hay *et al.* (1992) *Hum Antibod Hybridomas* 3:81-85; Huse *et al.* (1989) *Science* 246:1275-1281; McCafferty *et al.*, *Nature* (1990) 348:552-554; Griffiths *et al.* (1993) *EMBO J* 12:725-734; Hawkins *et al.* (1992) *J Mol Biol* 226:889-896; Clackson *et al.*
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(1991) *Nature* 352:624-628; Gram *et al.* (1992) *PNAS* 89:3576-3580; Garrad *et al.* (1991) *Bio/Technology* 9:1373-1377; Hoogenboom *et al.* (1991) *Nuc Acid Res* 19:4133-4137; and Barbas *et al.* (1991) *PNAS* 88:7978-7982, and PCT Publication No. WO 97/29131, the contents of each of which are incorporated herein by reference.

5 The recombinant antibody library may be from a subject immunized with IL-18, or a portion of IL-18 comprising the epitope of amino acids PLFEDMTDSDCRDNA (SEQ ID NO: 1). Alternatively, the recombinant antibody library may be from a naïve subject, *i.e.*, one who has not been immunized with IL-18, such as a human antibody library from a human subject who has not been immunized with human IL-18.

10 Antibodies of the invention are selected by screening the recombinant antibody library with the epitope of amino acids PLFEDMTDSDCRDNA (SEQ ID NO: 1) of human IL-18 to thereby select those antibodies that recognize this epitope. Methods for conducting such screening and selection are well known in the art, such as described in the references in the preceding paragraph.

15 To select antibodies of the invention having a particular binding affinity for hIL-18, the art-known method of surface plasmon resonance can be used. To select antibodies having a particular neutralizing activity for hIL-18, standard methods known in the art for assessing the inhibition of hIL-18 activity may be used. In addition, methods for immunizing mice that have been transgenically altered to encode a human
20 immunoglobulin repertoire thereby enabling the organism to express fully human antibodies in response to an immunogen, are known in the art (see, *e.g.*, U.S.P.N.s 5,877,397 and 6,150,584).

Uses of Anti-IL-18 Antibodies

25 Given their ability to bind to hIL-18, the anti-hIL-18 antibodies, or portions thereof, of the invention can be used to detect hIL-18 (*e.g.*, in a biological sample, such as serum or plasma), using a conventional immunoassay, such as an enzyme linked immunosorbent assays (ELISA), an radioimmunoassay (RIA) or tissue immunohistochemistry. The invention provides a method for detecting hIL-18 in a
30 biological sample comprising contacting a biological sample with an antibody, or antibody portion, of the invention and detecting either the antibody (or antibody portion) bound to hIL-18 or unbound antibody (or antibody portion), to thereby detect hIL-18 in the biological sample. The antibody is directly or indirectly labeled with a detectable substance to facilitate detection of the bound or unbound antibody. Suitable detectable
35 substances include various enzymes, prosthetic groups, fluorescent materials, luminescent materials and radioactive materials. Examples of suitable enzymes include horseradish peroxidase, alkaline phosphatase, β -galactosidase, or acetylcholinesterase; examples of suitable prosthetic group complexes include streptavidin/biotin and

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avidin/biotin; examples of suitable fluorescent materials include umbelliferone, fluorescein, fluorescein isothiocyanate, rhodamine, dichlorotriazinylamine fluorescein, dansyl chloride or phycoerythrin; an example of a luminescent material includes luminol; and examples of suitable radioactive material include ^{125}I , ^{131}I , ^{35}S , ^{32}P , ^{33}P , or ^3H .

Alternative to labeling the antibody, hIL-18 can be assayed in biological fluids by a competition immunoassay utilizing rhIL-18 standards labeled with a detectable substance and an unlabeled anti-hIL-18 antibody. In this assay, the biological sample, the labeled rhIL-18 standards and the anti-hIL-18 antibody are combined and the amount of labeled rhIL-18 standard bound to the unlabeled antibody is determined. The amount of hIL-18 in the biological sample is inversely proportional to the amount of labeled rhIL-18 standard bound to the anti-hIL-18 antibody.

The antibodies and antibody portions of the invention preferably are capable of neutralizing hIL-18 activity both *in vitro* and *in vivo*. Accordingly, such antibodies and antibody portions of the invention can be used to inhibit hIL-18 activity, *e.g.*, in a cell culture containing hIL-18, in human subjects or in other mammalian subjects having IL-18 with which an antibody of the invention cross-reacts. In one embodiment, the invention provides a method for inhibiting IL-18 activity comprising contacting IL-18 with an antibody or antibody portion of the invention such that IL-18 activity is inhibited. Preferably, the IL-18 is human IL-18. For example, in a cell culture containing, or suspected of containing hIL-18, an antibody or antibody portion of the invention can be added to the culture medium to inhibit hIL-18 activity in the culture.

In another embodiment, the invention provides a method for inhibiting IL-18 activity in a subject suffering from a disorder in which IL-18 activity is detrimental. The invention provides methods for inhibiting IL-18 activity in a subject suffering from such a disorder, which method comprises administering to the subject an antibody or antibody portion of the invention such that IL-18 activity in the subject is inhibited. Preferably, the IL-18 is human IL-18 and the subject is a human subject. Alternatively, the subject can be a mammal expressing an IL-18 with which an antibody of the invention cross-reacts. Still further the subject can be a mammal into which has been introduced hIL-18 (*e.g.*, by administration of hIL-18 or by expression of an hIL-18 transgene). An antibody of the invention can be administered to a human subject for therapeutic purposes. Moreover, an antibody of the invention can be administered to a non-human mammal expressing an IL-18 with which the antibody cross-reacts for veterinary purposes or as an animal model of human disease. Regarding the latter, such animal models may be useful for evaluating the therapeutic efficacy of antibodies of the invention (*e.g.*, testing of dosages and time courses of administration).

In particular, one animal model for modulating IL-18 activity in an animal uses NOD-SCID mice which are transplanted with human peripheral blood mononuclear cells. Then, two to four weeks after engraftment (as measured by human IgG titers in serum) the mice are injected with LPS (lipopolysaccharide). Four to six hours later LPS-induced human interferon-gamma serum titers are determined. The efficacy (potency) of anti-IL-18 antibodies (e.g., IL-18 neutralizing antibodies) is determined by injecting the antibodies (ip) one day prior to LPS challenge followed by monitoring the test animals for a reduction in interferon-gamma serum titers (a function of IL-18 *in vivo* activity) as compared to controls (see, e.g., Holmes et al., Hybridoma, 19:363367 (2000)).

As used herein, the term "a disorder in which IL-18 activity is detrimental" is intended to include diseases and other disorders in which the presence of IL-18 in a subject suffering from the disorder has been shown to be, or is suspected of being, either responsible for the pathophysiology of the disorder or a factor that contributes to a worsening of the disorder. Accordingly, a disorder in which IL-18 activity is detrimental is a disorder in which inhibition of IL-18 activity is expected to alleviate the symptoms and/or progression of the disorder. Such disorders may be evidenced, for example, by an increase in the concentration of IL-18 in a biological fluid of a subject suffering from the disorder (e.g., an increase in the concentration of IL-18 in serum, plasma, synovial fluid, etc. of the subject), which can be detected, for example, using an anti-IL-18 antibody as described above.

Non-limiting examples of disorders that can be treated with the antibodies of the invention include those disorders discussed in the section above pertaining to pharmaceutical compositions of the antibodies of the invention.

Other features of the invention will be apparent from the following examples which should not be construed as limiting.

EXEMPLIFICATION

In general, the practice of the present invention employs, unless otherwise indicated, conventional techniques of chemistry, molecular biology, recombinant DNA technology, PCR technology, immunology (especially, e.g., antibody technology), and any necessary cell culture or animal husbandry techniques, which are within the skill of the art and are explained fully in the literature. See, e.g., Sambrook, Fritsch and Maniatis, *Molecular Cloning: Cold Spring Harbor Laboratory Press* (1989); *DNA Cloning*, Vols. 1 and 2, (D.N. Glover, Ed. 1985); *Oligonucleotide Synthesis* (M.J. Gait, Ed. 1984); *PCR Handbook Current Protocols in Nucleic Acid Chemistry*, Beaucage, Ed. John Wiley & Sons (1999) (Editor); *Oxford Handbook of Nucleic Acid Structure*, Neidle, Ed., Oxford Univ Press (1999); *PCR Protocols: A Guide to Methods and*

- Applications, Innis et al., Academic Press (1990); *PCR Essential Techniques: Essential Techniques*, Burke, Ed., John Wiley & Son Ltd (1996); *The PCR Technique: RT-PCR*, Siebert, Ed., Eaton Pub. Co. (1998); *Quantitative PCR Protocols*, Kochanowski et al., Eds., Humana Press (1999); *Clinical Applications of PCR*, Lo, Ed., Humana Press
- 5 (1998); *Antibody Engineering Protocols (Methods in Molecular Biology)*, 510, Paul, S., Humana Pr (1996); *Antibody Engineering: A Practical Approach (Practical Approach Series, 169)*, McCafferty, Ed., Irl Pr (1996); *Antibodies: A Laboratory Manual*, Harlow et al., C.S.H.L. Press, Pub. (1999); *Current Protocols in Molecular Biology*, eds. Ausubel et al., John Wiley & Sons (1992); *Large-Scale Mammalian Cell Culture*
- 10 *Technology*, Lubiniecki, A., Ed., Marcel Dekker, Pub., (1990); and *Manipulating the Mouse Embryo*, Hogan et al., C.S.H.L. Press, Pub (1994).

Throughout the examples, unless otherwise indicated, the above materials and methods were used.

EXAMPLE 1

ISOLATION OF ANTI-IL-18 ANTIBODIES

- 15 Antibodies to hIL-18 were isolated by screening separate scFv phage display libraries prepared using human VL and VH cDNAs from mRNA derived from human B cells (e.g., tonsils and spleen). Construction of the library and methods for selection are
- 20 described in Vaughan et al. (1996) *Nature Biotech.* 14: 309-314.

- The libraries were screened using either full length human IL-18 (SEQ ID NO: 61), a peptide epitope of IL-18 (SEQ ID NOS: 1-3), or a panel of overlapping 15 amino acid peptides representing IL-18 (the epitope sequence of which is presented in Table 5; SEQ ID NOS: 31-60). IL-18 specific antibodies were selected by coating the antigen
- 25 onto immunotubes using standard procedures (Marks et al., (1991) *J. Mol. Biol.* 222: 581-597). The scFv libraries were screened using either IL-18, a peptide epitope of IL-18, or an IL-18 peptide panel to generate a significant number of IL-18 specific binders. Several different clonotypes were selected, determined by restriction enzyme digestion patterns, and confirmed by DNA sequencing.

- 30 In order to identify IL-18 antibodies which preferentially bind either full length IL-18 or a representative peptide thereof, the supernatant containing scFv was titrated on biotin-captured IL-18 in an ELISA and binding characteristics were determined.

- Two anti-IL-18 single chain antibodies were obtained, one termed 2E1, independently isolated using a peptide epitope and the peptide panel, and a second anti-
- 35 IL-18 antibody termed LT28, isolated using full length IL-18. These parent anti-IL-18 antibodies were selected for further study and modification.

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EXAMPLE 2**AFFINITY MATURATION OF AN ANTI-IL-18 ANTIBODIES**

A single chain Fv version of antibody 2E1 having an identified IL-18 binding activity and the heavy chain and light chain sequence shown in Table 6 was further modified for improved neutralization of IL-18 activity.

Table 6. Sequence of Single-Chain Anti-IL-18 Antibody 2E1

2E1 Heavy Chain (SEQ ID NO: 18)	
CDR1 (SEQ ID NO: 9) QVQLVQSGAEVKKPGASMKVSCKTSGYTF <u>SGYYIH</u> WVRQAHGQGFWEI	
CDR2 (SEQ ID NO: 10) <u>GRLNPTTGDANFAEK</u> FG	CDR3 (SEQ ID NO: 11) GRVALTRDTSISTAYLQLDSLKSDDTAVYYCAG <u>KEGA</u> WGQGLVTVSS
2E1 Light Chain (SEQ ID NO: 19)	
CDR1 (SEQ ID NO: 12) SSELTPDPAVSVALGQTVRITC <u>QGDSLRFY</u> PNWYQQKPGQAPVLVIY	CDR2 (SEQ ID NO: 13) <u>GKNNRPS</u>
CDR3 (SEQ ID NO: 14) GIPDRFSGSGSGNTGSLTITGAQAEDEADYYC <u>GSRDSSGIHVV</u> FGGGTKVTVLG	

The anti-IL-18 antibody 2E1 was independently selected using an IL-18 peptide and sequential, overlapping, peptide panel representative of IL-18 (see Table 6).

The specific amino acid residues of the heavy chain variable region selected for mutagenesis are summarized in Table 7. In particular, with respect to the heavy chain region, individual amino acid substitutions were tested at positions H30, H31, H32, H33, and H35 of CDR1, positions H52, H52a, H53, H54, H56, and H58 of CDR2, and H95, H96, H97, and H98 of CDR3.

With regards to light chain amino acid residues selected for mutagenesis, individual amino acid substitutions were tested at positions L30, L31, L32, and L34 of CDR1, positions L50, L52, L53, and L55 of CDR2 and positions L89, L90, L91, L92, L93, L94, L95, L95a, L95b, L96, and L97 of CDR 3.

Table 7. Heavy Chain Amino Acid Substitutions Introduced Into 2E1

Heavy Chain Mutations	
CDR / Kabat Position	substituted residue
CDR1	
H30	A, R, N, D, C, G, H, I, F, P, S, or V
H31	A, C, H, S, T, or Y
H32	R, N, C, H, P, S, or T
H33	N, D, C, Q, H, L, M, F, S, or V
H35	N, D, L, or F
CDR2	
H52	T
H52a	R, Q, L, S, T, or W
H53	A, R, N, L, P, S, or Y
H54	A, R, N, D, Q, L, K, M, P, S, or Y
H56	A, R, N, C, G, H, I, L, or F
H58	A, R, Q, E, H, I, L, K, M, F, S, T, Y, P, S, T, W, Y, or V
CDR3	
H95	A, R, E, Q, S, Y, V, H, P, W, or C
H96	A, R, Q, S, Y, V, H, P, W, or C
H97	A, R, E, Q, S, Y, V, H, P, W, or C
H98	R, E, Q, S, Y, V, H, P, W, or C

Table 8. Light Chain Amino Acid Substitutions Introduced Into 2E1

5

Light Chain Mutations	
CDR / Kabat Position	substituted residue
CDR1	
L30	N, D, C, G, I, L, S, W, or Y
L31	R, N, D, C, G, H, I, L, P, S, T, or Y
L32	R, N, D, E, G, I, L, P, S, T, or V
L34	A, R, N, D, E, H, I, L, K, M, F, P, S, T, Y, or V
CDR2	
L50	A, N, I, L, F, P, S, W, Y, or V
L52	A, R, D, E, H, I, L, M, F, P, S, T, or V
L53	A, R, C, I, L, K, M, P, S, or T
L55	A, R, N, D, C, G, H, I, L, S, T, or Y
CDR3	
L89	A, R, E, Q, S, Y, V, H, P, W, or C
L90	A, R, E, Q, Y, V, H, P, W, or C
L91	R, E, Q, S, Y, V, H, P, W, or C
L92	A, R, E, Q, S, Y, V, H, P, W, or C
L93	A, R, E, Q, Y, V, H, P, W, or C
L94	A, R, E, Q, Y, V, H, P, W, or C
L95	A, R, E, Q, S, Y, V, H, P, W, or C
L95a	A, R, E, Q, S, Y, V, H, P, W, or C
L95b	A, R, E, Q, S, Y, V, P, W, or C
L96	A, R, E, Q, S, Y, H, P, W, or C
L97	A, R, E, Q, S, Y, H, P, W, or C

Substitutions were introduced using standard techniques (e.g., as described in Taylor et al., *Nucleic Acids Res.* 13: 8764-8758 (1985); Nakamaye et al., *Nucleic Acids Res.* 14: 9679-9698 (1986); and Olsen et al., *Methods in Enzymology*, 217: 189 (1993)). In brief, oligonucleotides degenerate for a given codon were synthesized for each of the positions to be mutagenized. A single-stranded DNA template was prepared from the original plasmid containing a single-chain Fv version of the antibody 2E1 gene. The nucleic acid sequence of the parent 2E1 antibody heavy and light chain is provided in SEQ ID NOS: 62 and 64. The mutant oligonucleotides were then used to create a complementary DNA strand and eventually a double-stranded plasmid, thus incorporating the degeneracy or the different mutations in a given codon of the antibody. In particular, the CDR3 region of the heavy and light chain of 2E1 was altered using the QuikChange Kit (Stratagene) according to the manufactures instruction.

A representative number of clones were then sequenced from each mutagenesis reaction (i.e., 7 to 36 clones) and those representing a change from the parent 2E1 single chain antibody sequence were expressed in bacteria and purified for further *in vitro* and *in vivo* testing as described infra.

In another screen using a full length IL-18 ligand, a second anti-IL-18 antibody was identified and selected for further improvement using affinity maturation. In particular, using the techniques described above, the LT28 antibody having the heavy chain and light chain sequence shown in Table 9 (and nucleic sequence provided in SEQ ID NOS: 66 and 68) was further modified.

Table 9. Sequence of Single-Chain Anti-IL-18 Antibody LT28

LT28 Heavy Chain (SEQ ID NO: 28)	
CDR1 (SEQ ID NO: 20)	CDR2 (SEQ ID NO: 21)
LVQPGGSLRLSCAASGFTFS	<u>SYAMS</u> WVRQAPGKLEWVS <u>AI</u> <u>SGSGGSTYYADSVKG</u>
CDR3 (SEQ ID NO: 22)	
RFTISRDN SKNTLYLQMNSLRAEDTAVYYCAR <u>DDDDYDFDY</u> WGRGTMVTVSS	
LT28 Light Chain (SEQ ID NO: 29)	
CDR1 (SEQ ID NO: 23)	CDR2 (SEQ ID NO: 24)
QSVLTQPPSASGTPGQRTIS	<u>CGSSSNIGINAVN</u> WYQQLPGTAPKLLIY <u>GNDQRPS</u>
CDR3 (SEQ ID NO: 25)	
GVPDFRFGSKSGTSASLAISGLQSEDEADYYC <u>AAWDDSLSGPFV</u> FGGGTKLTVLG	

With respect to the heavy chain region, amino acid substitutions were introduced at positions H31, H32, H33, and H35 of CDR1, positions H50, H51, H52, H52a, H53, H54, H56, and H58 of CDR2, and H95, H96, H97, H98, H99, H100, H100a, H101, and
 5 H102 of CDR3.

With regards to light chain residues selected for mutation, amino acid substitutions were introduced at positions L30, L31, L32, L34 of CDR1, positions L50, L52, L53, L55 of CDR2 and positions L89, L90, L91, L92, L93, L94, L95, L95a, L95b, L96, L97.
 10

Table 10. Heavy Chain Amino Acid Substitutions Introduced into LT28

Heavy Chain Mutations	
CDR / Kabat Position	substituted residue
CDR1	
H31	A, R, E, Q, S, Y, V, H, P, W, or C
H32	A, R, E, Q, S, Y, V, H, P, W, or C
H33	A, R, E, Q, S, Y, V, H, P, W, or C
H35	A, R, E, Q, S, Y, V, H, P, W, or C
CDR2	
H50	A, R, E, Q, S, Y, V, H, P, W, or C
H51	A, R, E, Q, S, Y, V, H, P, W, or C
H52	A, R, E, Q, S, Y, V, H, P, W, or C
H52a	A, R, E, Q, S, Y, V, H, P, W, or C
H53	A, R, E, Q, S, Y, V, H, P, W, or C
H54	A, R, E, Q, S, Y, V, H, P, W, or C
H56	A, R, E, Q, S, Y, V, H, P, W, or C
H58	A, R, E, Q, S, Y, V, H, P, W, C
CDR3	
H95	A, R, E, Q, S, Y, V, H, P, W, or C
H96	A, R, E, Q, S, Y, V, H, P, W, or C
H97	A, R, E, Q, S, Y, V, H, P, W, or C
H98	A, R, E, Q, S, Y, V, H, P, W, or C
H99	A, R, E, Q, S, Y, V, H, P, W, or C
H100	A, R, E, Q, S, Y, V, H, P, W, or C
H100a	A, R, E, Q, S, Y, V, H, P, W, or C
H101	A, R, E, Q, S, Y, V, H, P, W, or C
H102	A, R, E, Q, S, Y, V, H, P, W, or C

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Table 11. Light Chain Amino Acid Substitutions Introduced into LT28

Light Chain Mutations	
CDR / Position	substituted residue
CDR1	
L30	A, R, E, Q, S, Y, V, H, P, W, C
L31	A, R, E, Q, S, Y, V, H, P, W, C
L32	R, E, Q, S, Y, V, H, P, W, C, G
L34	A, R, E, Q, S, Y, V, H, P, W, C
CDR2	
L50	A, R, E, Q, S, Y, V, H, P, C
L52	A, R, E, S, Y, V, H, P, W, C
L53	A, R, E, S, Y, V, H, P, W, C, N
L55	A, E, Q, S, Y, V, H, P, W, C
CDR3	
L89	A, R, E, Q, S, Y, V, H, P, W, C
L90	A, R, E, Q, S, Y, V, H, P, W, C
L91	A, R, E, Q, S, Y, V, H, P, W, C
L92	A, R, E, Q, S, Y, V, H, P, W, C
L93	A, R, E, Q, S, Y, V, H, P, W, C
L94	A, R, E, Q, S, Y, V, H, P, W, C
L95	A, R, E, Q, S, Y, V, H, P, W, C
L95a	A, R, E, Q, S, Y, V, H, P, W, C
L95b	A, R, E, Q, S, Y, V, H, P, W, C
L96	A, R, E, Q, S, Y, V, H, P, W, C
L97	A, R, E, Q, S, Y, V, H, P, W, C

- 5 Substitutions were introduced as described above. A representative number of clones were then sequenced from each mutagenesis reaction and those representing a change from the parent LT28 single chain antibody sequence were expressed in bacteria and purified for further testing as described below.

10

EXAMPLE 3**BINDING ACTIVITY OF HUMAN ANTIBODIES TO IL-18**

- Real-time binding interactions between ligand (biotinylated recombinant human IL-18 (rhIL-18) immobilized on a biosensor matrix) and analyte (antibodies in solution) were measured by surface plasmon resonance (SPR) using the BIAcore system
- 15 (Pharmacia Biosensor, Piscataway, NJ). The system utilizes the optical properties of SPR to detect alterations in protein concentrations within a dextran biosensor matrix. Proteins are covalently bound to the dextran matrix at known concentrations. Antibodies are injected through the dextran matrix and specific binding between injected antibodies and immobilized ligand results in an increased matrix protein concentration
- 20 and resultant change in the SPR signal. These changes in SPR signal are recorded as

resonance units (RU) and are displayed with respect to time along the y-axis of a sensorgram.

To facilitate immobilization of biotinylated rhIL-18 on the biosensor matrix, streptavidin is covalently linked via free amine groups to the dextran matrix by first activating carboxyl groups on the matrix with 100 mM N-hydroxysuccinimide (NHS) and 400 mM N-ethyl-N'-(3-diethylaminopropyl) carbodiimide hydrochloride (EDC). Next, streptavidin is injected across the activated matrix. Thirty-five microliters of streptavidin (25 µg/ml), diluted in sodium acetate, pH 4.5, is injected across the activated biosensor and free amines on the protein are bound directly to the activated carboxyl groups. Unreacted matrix EDC-esters are deactivated by an injection of 1 M ethanolamine. Streptavidin-coupled biosensor chips also are commercially available (Pharmacia BR-1000-16, Pharmacia Biosensor, Piscataway, NJ).

Biotinylated rhIL-18 was prepared by first dissolving 5.0 mg of biotin (D-biotinyl-ε-aminocaproic acid N-hydroxysuccinimide ester; Boehringer Mannheim Cat. No. 1008 960) in 500 µl dimethylsulfoxide to make a 10 mg/ml solution. Ten microliters of biotin was added per ml of rhIL-18 (at 2.65 mg/ml) for a 2:1 molar ratio of biotin to rhIL-18. The reaction was mixed gently and incubated for two hours at room temperature in the dark. A PD-10 column, Sephadex G-25M (Pharmacia Catalog No. 17-0851-01) was equilibrated with 25 ml of cold PBS and loaded with 2 ml of rhIL-18-biotin per column. The column was eluted with 10 x 1 ml cold PBS. Fractions were collected and read at OD280 (1.0 OD = 1.25 mg/ml). The appropriate fractions were pooled and stored at -80° C until use.

Biotinylated rhIL-18 to be immobilized on the matrix via streptavidin was diluted in PBS running buffer (Gibco Cat. No. 14190-144, Gibco BRL, Grand Island, NY) supplemented with 0.05% (BIAcore) surfactant P20 (Pharmacia BR-1000-54, Pharmacia Biosensor, Piscataway, NJ). To determine the capacity of rhIL-18-specific antibodies to bind immobilized rhIL-18, a binding assay was conducted as follows. Aliquots of biotinylated rhIL-18 (25 nM; 10 µl aliquots) were injected through the streptavidin-coupled dextran matrix at a flow rate of 5 µl/min. Before injection of the protein and immediately afterward, PBS buffer alone flowed through each flow cell. The net difference in signal between baseline and approximately 30 sec. after completion of biotinylated rhIL-18 injection was taken to represent the binding value. Direct rhIL-18-specific antibody binding to immobilized biotinylated rhIL-18 was measured. Antibodies (20 µg/ml) were diluted in PBS running buffer and 25 µl aliquots were injected through the immobilized protein matrices at a flow rate of 5 µl/min. Prior to injection of antibody, and immediately afterwards, PBS buffer alone flowed through each flow cell. The net difference in baseline signal and signal after completion of antibody injection was taken to represent the binding value of the particular sample.

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Biosensor matrices were regenerated using 100 mM HCl before injection of the next sample. To determine the off rate (K_{off}), on rate (K_{on}), association rate (K_a) and dissociation rate (K_d) constants, BIAcore kinetic evaluation software (version 2.1) was used.

- 5 Representative results of improved candidate anti-IL-18 antibodies binding to biotinylated rhIL-18, as compared to the parent antibodies 2E1 and LT28 (and murine controls), are shown below in Table 12. For comparison, IC50 values from the cell-based neutralization assay are also included and these are described in Example 4. All clones were prepared as single-chain Fv antibodies for testing using Biacore analysis
- 10 and the cell-based assay described below. Parental clones listed comprise an unmutated parental heavy and light chain, whereas single chain mutants contain one parental chain and one mutated chain where the mutated chain is indicated as being either heavy (H) or light (L) followed by the Kabat position and nature of the amino acid substitution.

15 **Table 12. Binding of Anti-IL-18 Antibodies Derived From 2E1 and LT28**

Antibody Clone	On-rates ($M^{-1}s^{-1}$)	Off-rates (s^{-1})	Kd (M)	IC50 Value*
2E1 parent and mutants				
2E1 (parent) ScFv	2.6E+3	6.42E-03	1.5E-07	3.3E-8M
2E1 (parent) IgG				9.0E-10M
L34S		1.69E-04		1.5E-8M
H53R		2.34E-03		2.5E-8M
H53Y		-		1.5E-8M
H58Q		-		1.6E-8M
L34S + H53R (2E1RS)	2.7E+03	6.82E-05	2.3E-08	3.0E-09M
L34S + H58Q		-		1.5E-8M
L34S + H53Y		5.28E-05		6.7E-9M
H53R + H58Q		-		1.2E-8M
H53Y + H58Q		-		1.2E-8M
L34S + H53R + H58Q		6.18E-05		2.8E-9M
L34S + H53Y + H58Q		-		8.0E-9M
L90C				4x
L93C				2-4x
L94P, Q, or R				2-4x
L95R, Y				3-8x
L95bE, W				2-4x
LT28 parent and mutants				
LT28 (parent)	1.3E+04	4.8E-04	3.9E-08	9.0E-8M
H54Q				2-3x
H58W				2-3x
1				
125-2H	1.7E+05	1.1E-04	6.2E-10	2.E-10M
318-M	1.2E+04	1.1E-04	9.6E-09	4.0E-9M

* Some values presented as fold improvement compared to the parent.

EXAMPLE 4

NEUTALIZING ACTIVITY OF ANTI-IL-18 ANTIBODIES

To examine the neutralizing activity of the anti-human IL-18 antibodies of the invention, an art recognized assay for monitoring IL-18 activity was used.

5 Briefly, the assay employs KG1 cells (ATCC #CCL-246, myelogenous leukemia bone marrow cells) which were cultured according to standard techniques (*e.g.*, using RPMI 1640 Culture Medium Gibco #21870-076; (supplemented with 10% Fetal Bovine Serum (BioWhittaker #14-501F); 2mM L-glutamine (Gibco #25030-081); 50units/ml Penicillin, 50ug/ml Streptomycin (Gibco #15070-063); and .075% Sodium Bicarbonate).

10 To test for IL-18 neutralization, 3×10^5 KG-1 cells stimulated with 20ng/ml hTNF-alpha (Lot# 19130132) was incubated with 50ul of anti-IL-18 antibody (4 x Conc.) and 50ul of IL-18 (4xConc.=8ng/ml) and incubated for 37°C for 1hr or 16-20 hrs. To determine the amount of IL-18 neutralization that occurred as a function of induced hIFN-gamma production, an ELISA was performed using commercially available Elisa
15 Kits (R & D #DIF00/Endogen #EH-IFNG), according to the manufacturers instructions, and hIFN-gamma production was calculated (pg/ml) off a standard curve.

In all, four mutant antibodies, *i.e.*, the 2E1 derived L34S, H53R, H53Y, and H58Q, were shown to have greater IL-18 neutralization potency than the parent 2E1 antibody (see Table 12). The improvements in IC50 values using the KG-1 assay were
20 in the range of 2 to 5 fold, and similar improved binding results were determined using BIAcore analysis.

Various mutation combination clones were also prepared and tested, and this data is summarized in the Table 12. The best combination clone L34S-H53R showed a greater than ten fold improvement over the parent antibody 2E1 in both the KG-1 cell-
25 based assay and using the BIAcore analysis. The resulting antibody was designated the name of 2E1RS.

Several other mutant clones of 2E1 showed an improvement in potency, *i.e.*, IL-18 neutralization, as determined using the KG-1 assay. The mutant L95Y offered 5 to 8 fold better IC50 values than the parent 2E1 antibody. Several other mutants offered a 2
30 to 3 fold improvement and they are 2E1 mutants H96A, H96Q, H96S, H98S, L90C, L90W, L93C, L94P, L94P, L94Q, L94R, L94W, L95R, L95aA, L95aH, L95aP, L95aR, L95aW, L95bE, L95bW, L95bY, L97C, and L97E.

The binding of 2E1 in the form of an ScFv antibody or IgG antibody was also compared (see Fig. 5).

35 Still further, two mutants derived from the LT28 parent should improved IL-18 neutralization activity compared to the parent antibody.

These results demonstrate that fully human IL-18 neutralizing antibodies can be obtained using the methods and compositions of the invention.

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EQUIVALENTS

Those skilled in the art will recognize, or be able to ascertain using no more than routine experimentation, many equivalents to the specific embodiments of the invention

- 5 described herein. Such equivalents are intended to be encompassed by the following claims.

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